

# Monoclonal Antibody Synthesis Process – mAb-X

From the cell bank to the final product

**Goal :** Deliver a safe, effective IgG1 drug product (mAb-X) from qualified cell bank

**Upstream  
(USP)**

**Downstream  
(DSP)**

**Filling and  
Finishing**

# Description of Antibody Quality Attributes

## Common product quality attributes for therapeutic antibodies

### Disulfide bonds

Mis-paired / scrambled  
disulfides

### Cyclization

N-terminal pyroglutamate

### Integrity

Fragmentation, half-  
antibodies, free light chain

### C-terminal clipping

Lysine truncation

### Glycosylation (Fc)

GOF/G1F/G2F, afucosylation,  
high-mannose, sialylation

### Glycation

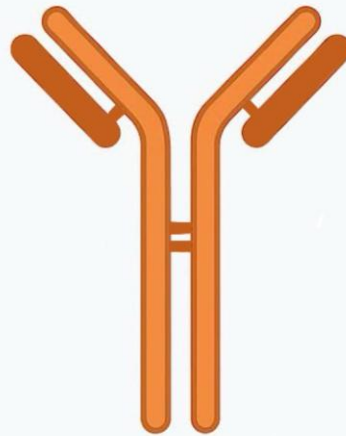
Non-enzymatic (reducing  
sugars)

### Chemical modifications

Methionine oxidation  
isomerisation

### Higher-order structure

Aggregation, dimerization



# Quality Attributes of the Final Drug Product

## Formulation :

### Final Formulation of mAb-X

- IgG1 Antibody: 10 mg/mL
- Phosphate buffer: 10 mM
- NaCl : 150 mM
- Polysorbate 80 : 0.01%
- pH:  $6.0 \pm 0.2$
- Osmolality:  $\sim 290$  mOsm/kg

## Drug Product Packaging :



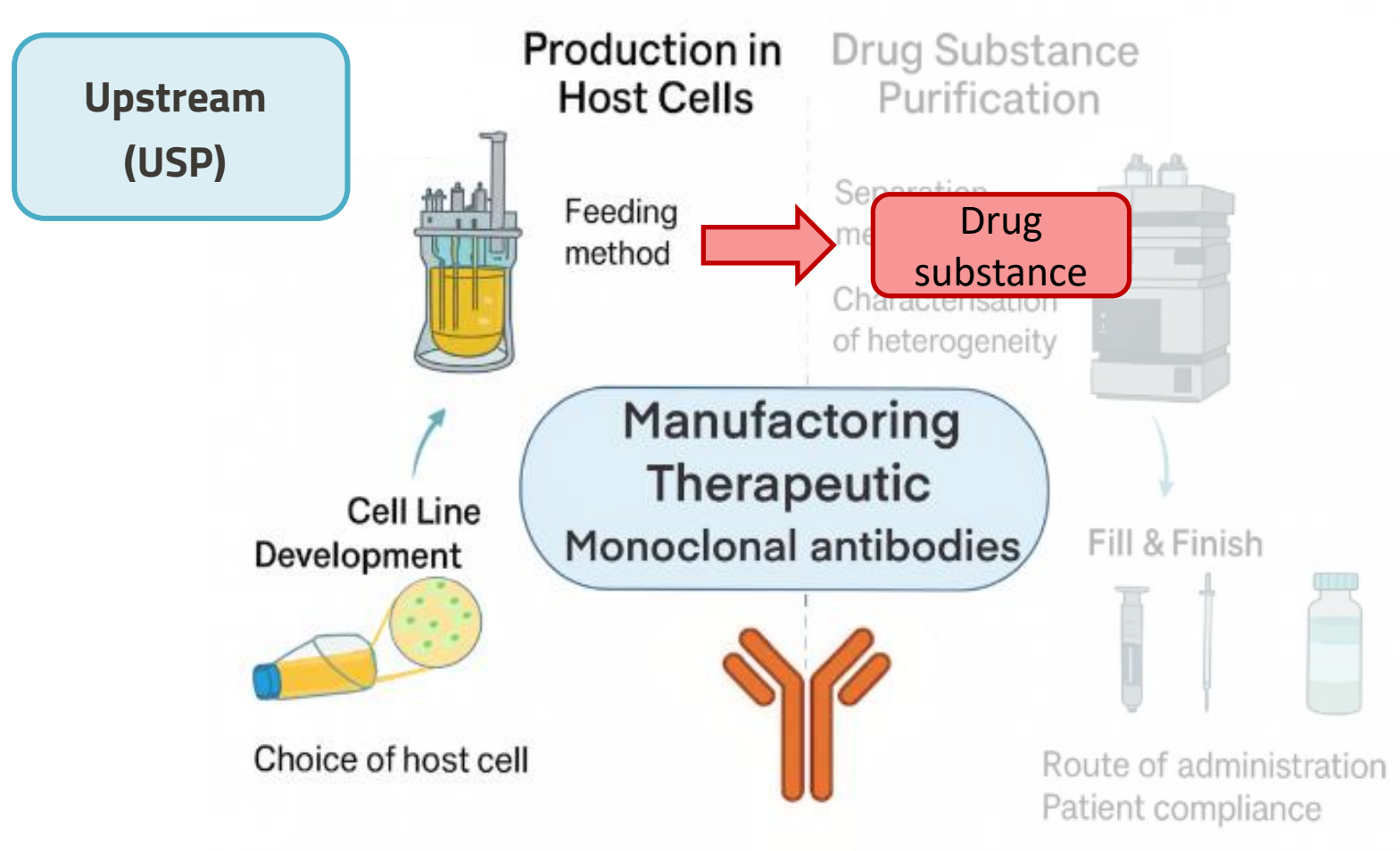
## Stability

24 month at  $2-8^{\circ}\text{C}$  ,  
short-term: 30 days at  $25^{\circ}\text{C}$

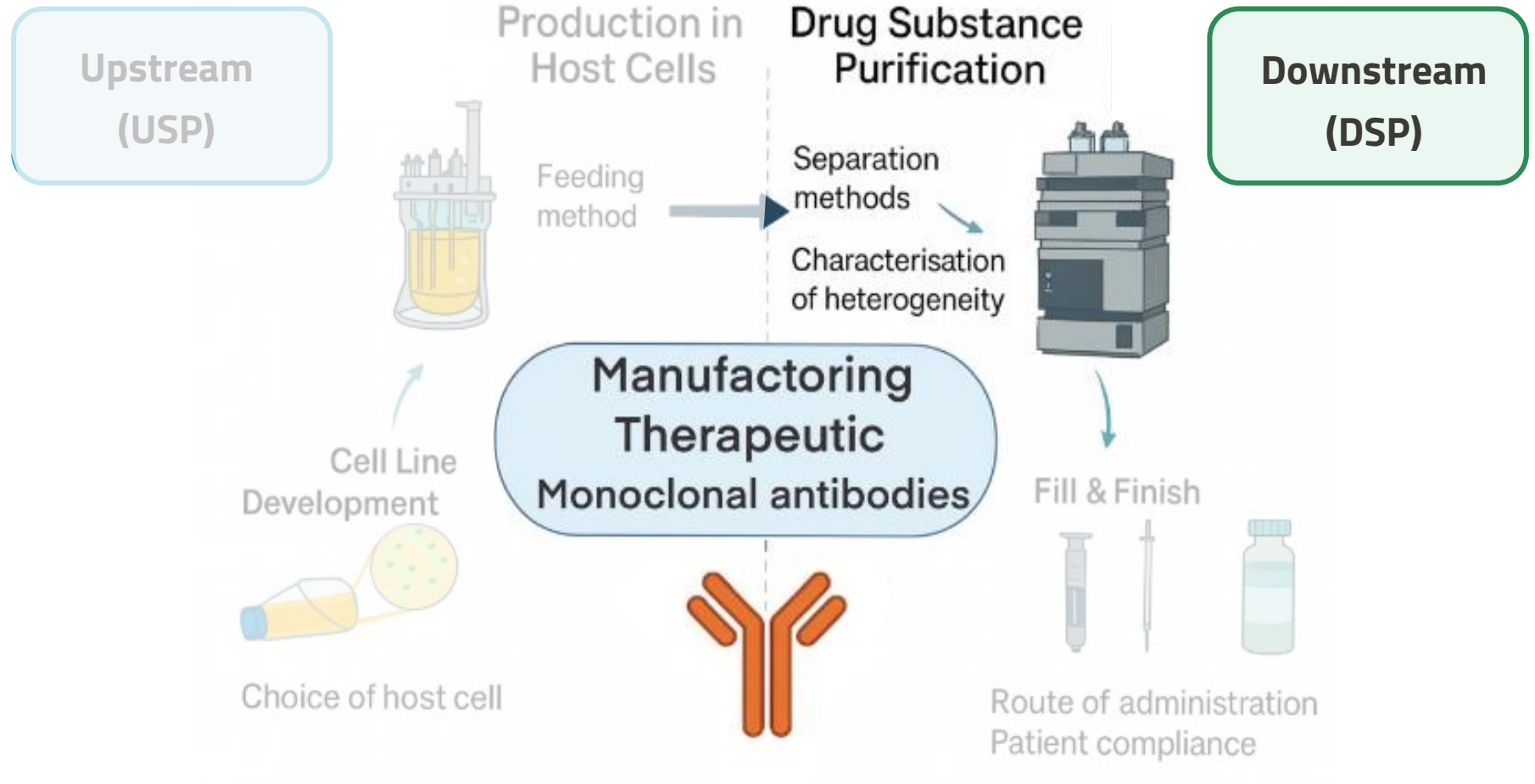
## Release specifications:

Purity  $\geq 98\%$        $\text{IC}_{50} \leq 15$  ng/mL  
Aggregates  $< 1\%$       HCP  $< 5$  ppm

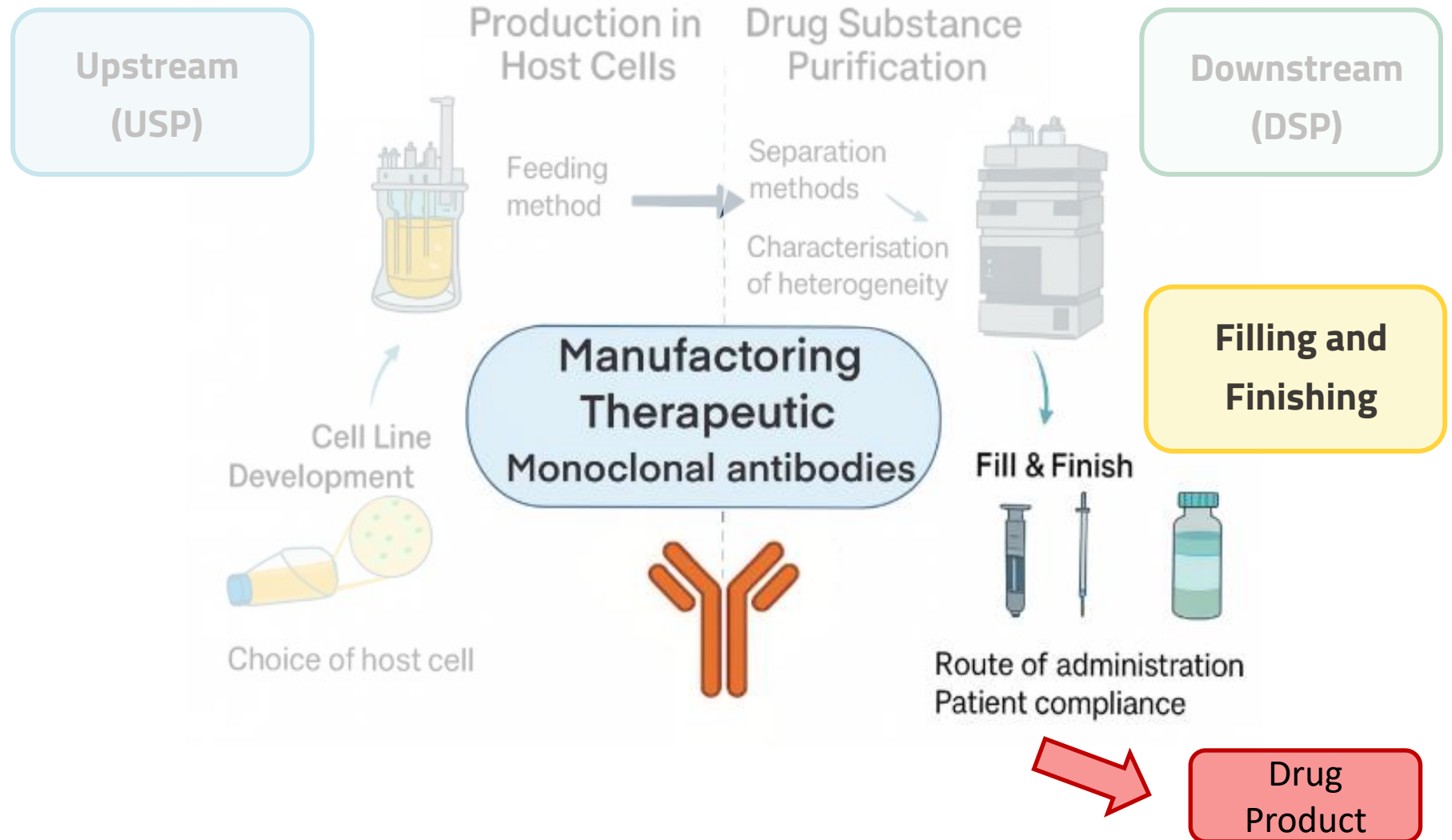
# Global Process Flow



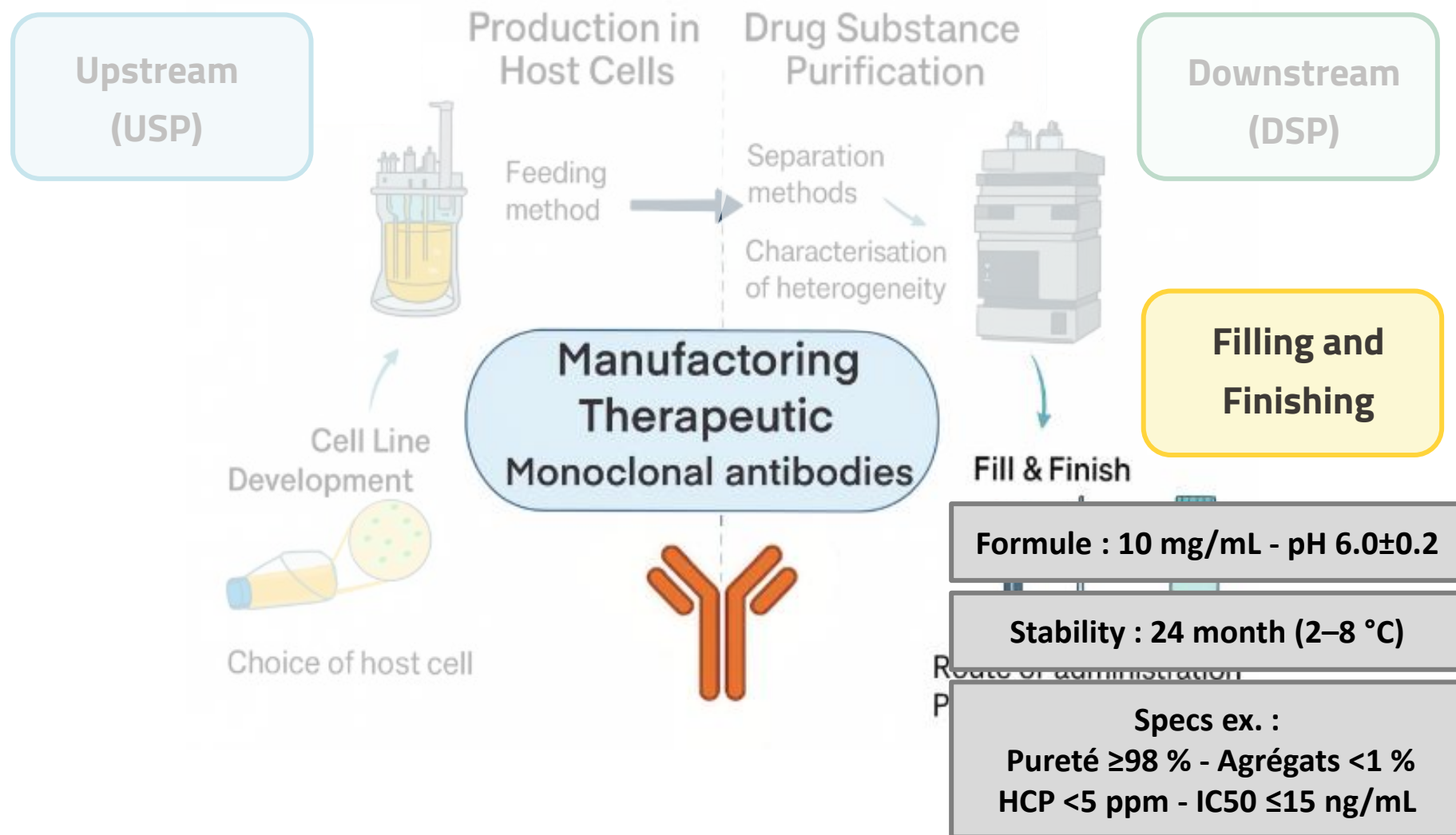
# Global Process Flow



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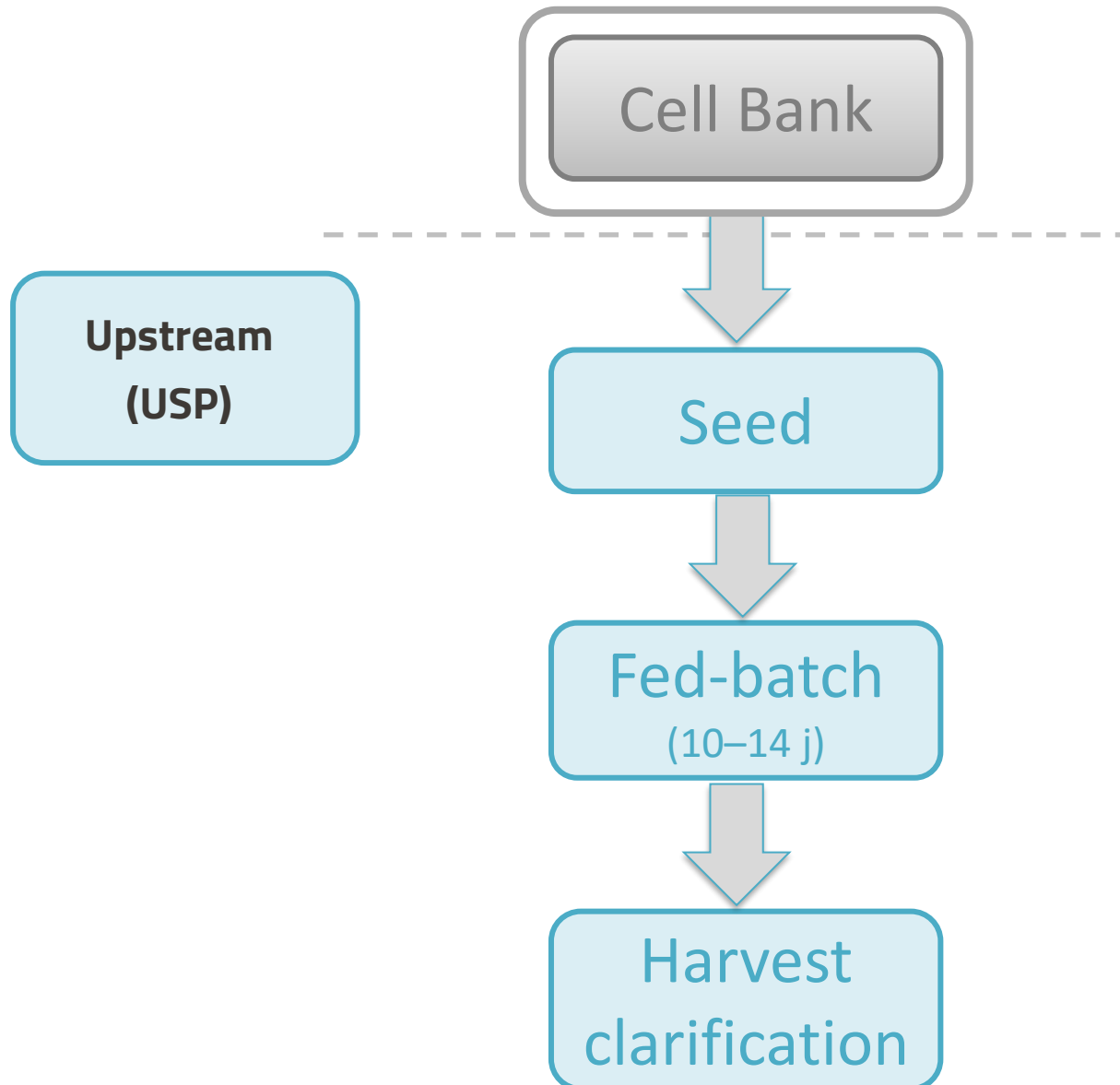


# Global Process Flow



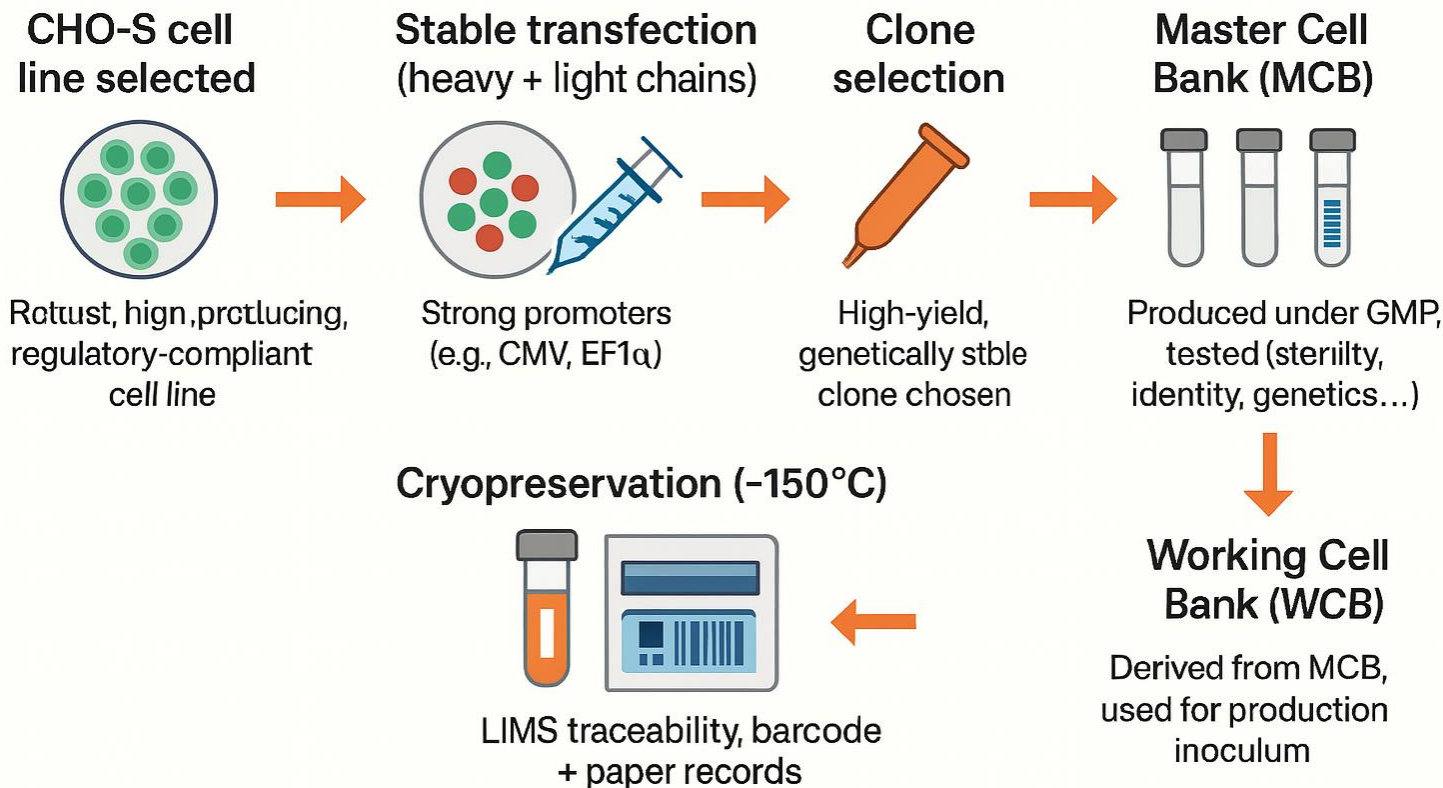


# Upstream Process Flow



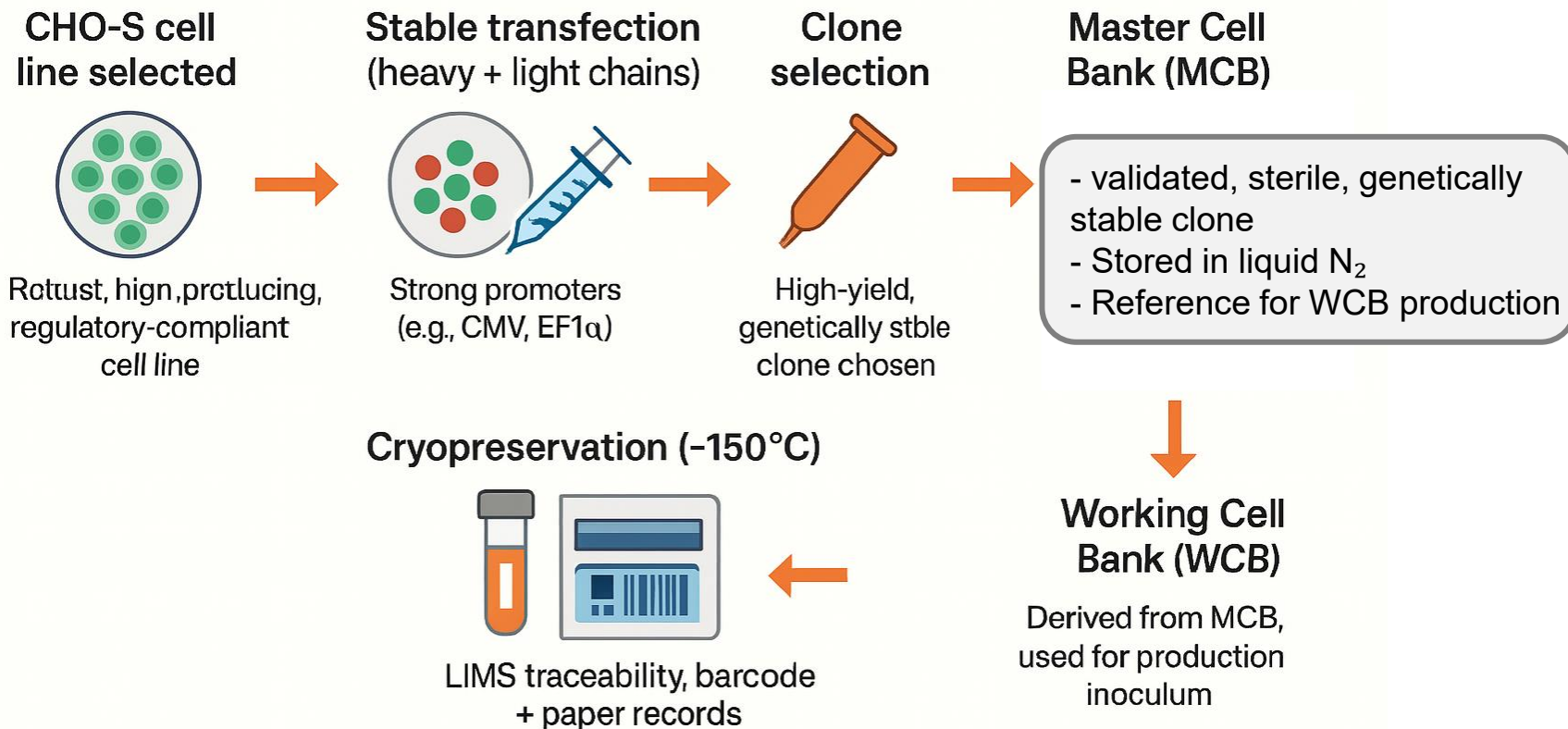
# Before UPSTREAM Process

## Cell Line Development and Banking Workflow



# Before UPSTREAM Process

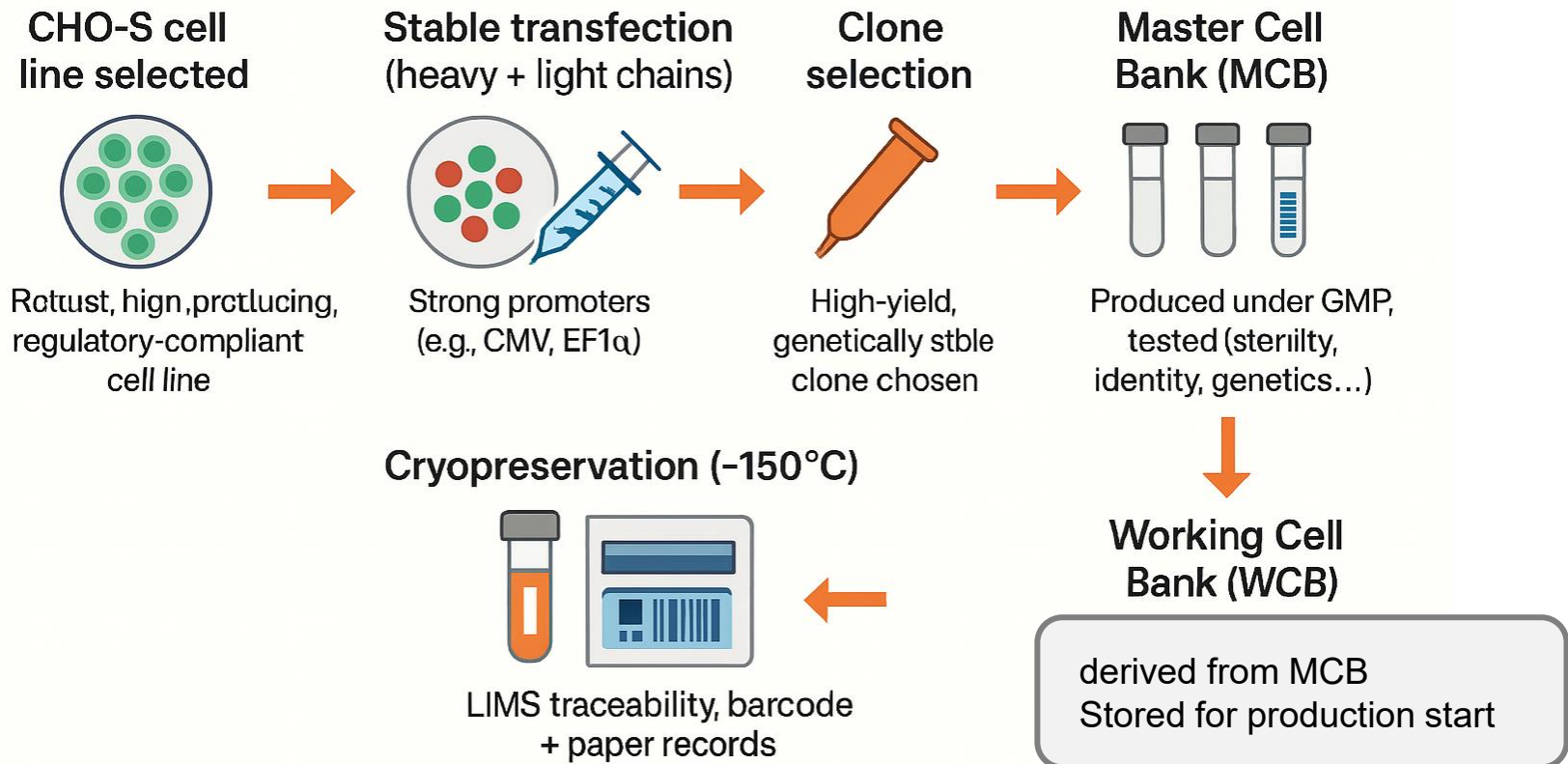
## Cell Line Development and Banking Workflow



**Tracked in the LIMS:** unique ID, location and time-stamped history of cryotubes, **Quality Control**, and status

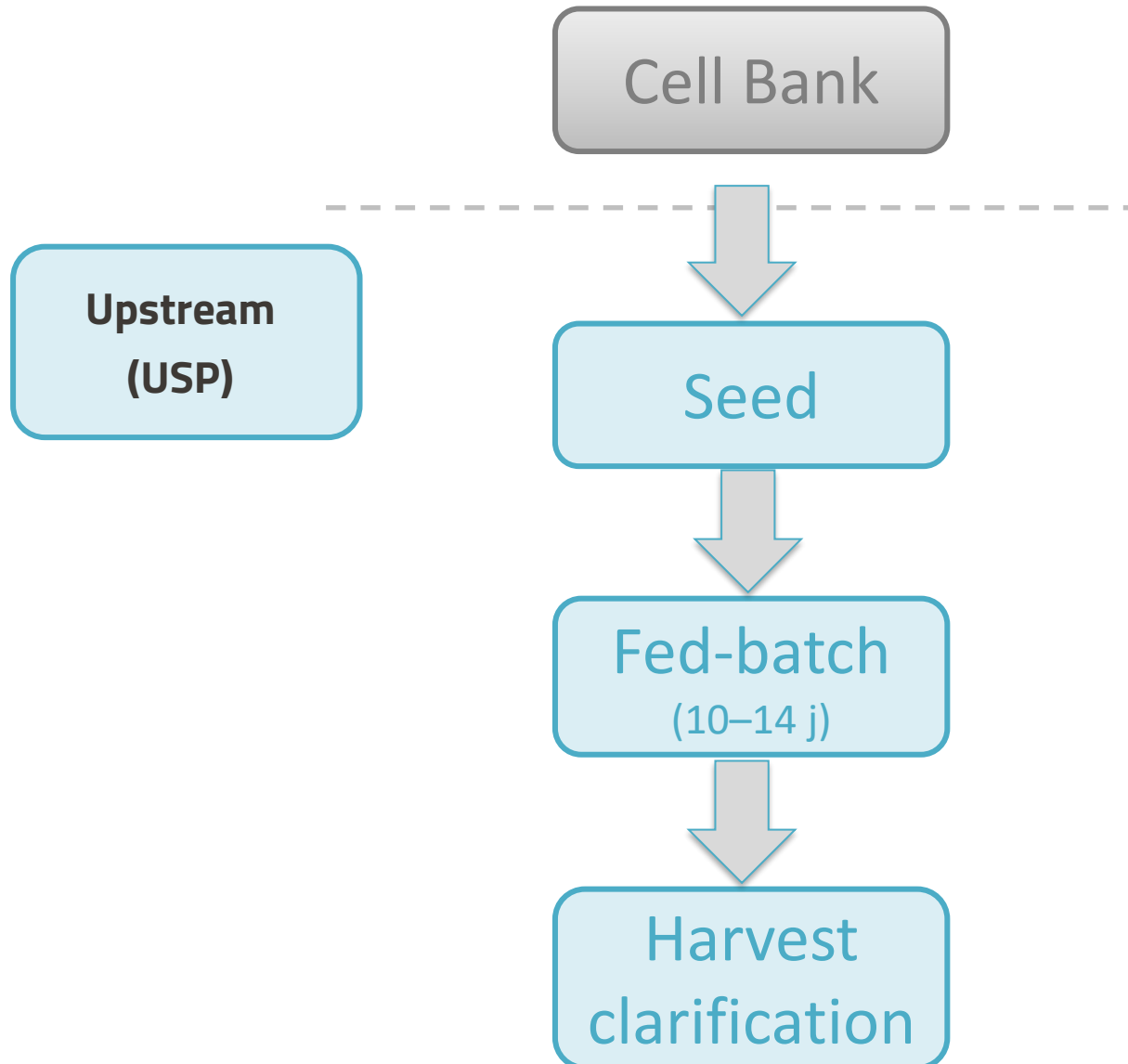
# Before UPSTREAM Process

## Cell Line Development and Banking Workflow



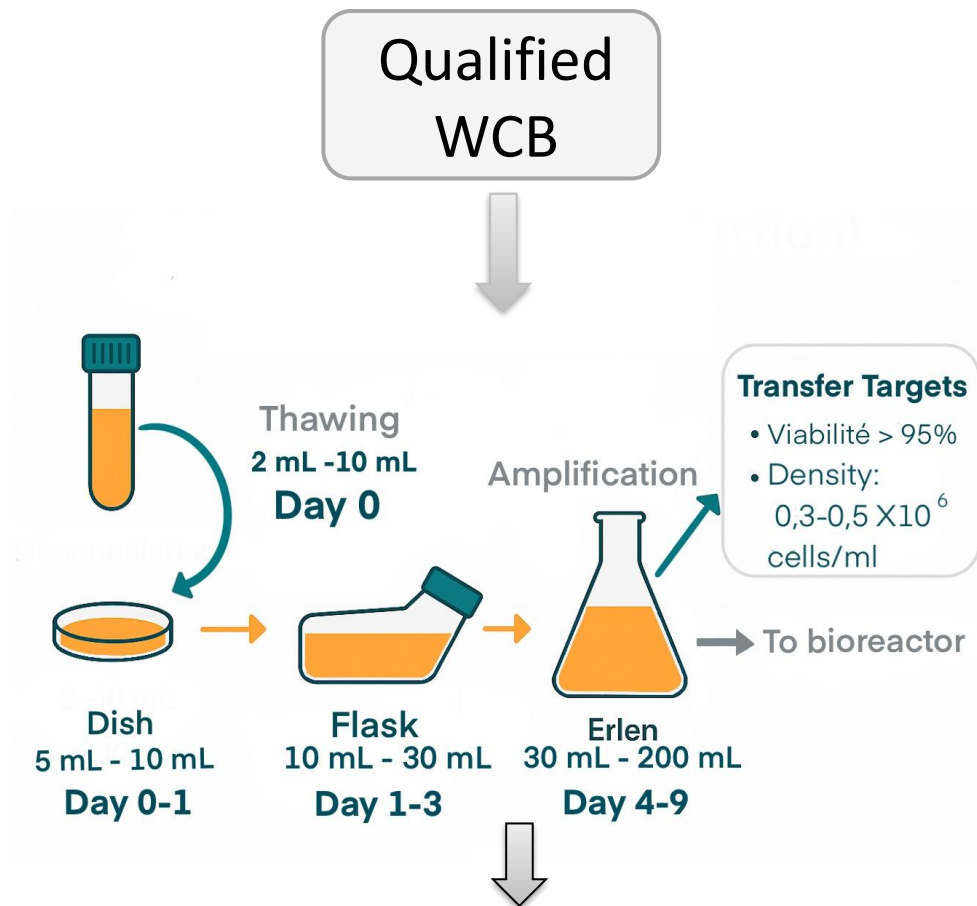
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# Upstream Process Flow



# Cell Bank Thawing and Pre-culture

Preparation of a viable and stable  
inoculum from the Working cell bank



**Outputs :** Viable inoculum for seed train

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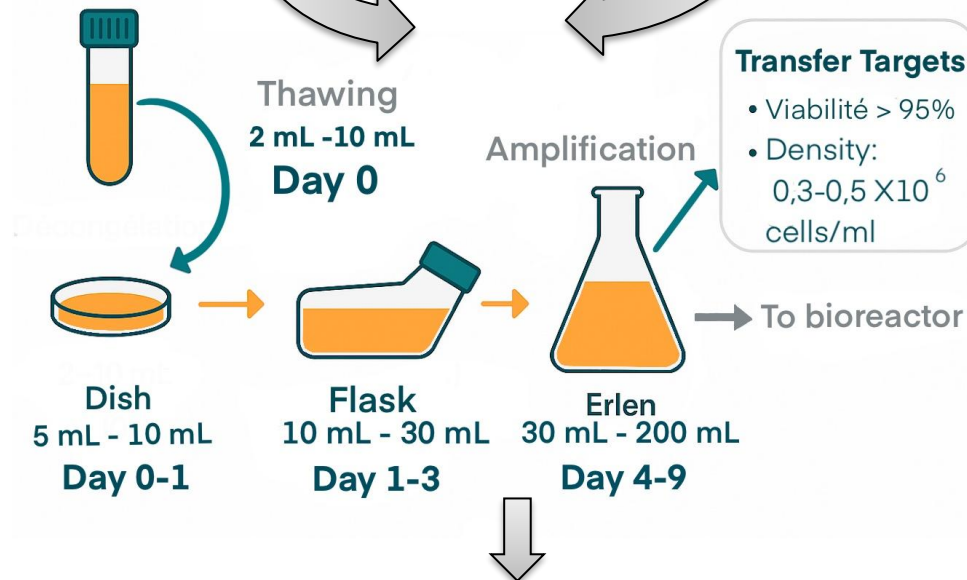
MA

Animal component–  
free medium  
Feed and supplement

Qualified  
WCB

PP

post-thaw  
temp, CO<sub>2</sub>, pH, Feed  
schedule



**Outputs :** Viable inoculum for seed train  
IPCs : viability, growth rate, morphology

# Cell Bank Thawing and Pre-culture

Preparation of a viable and stable  
inoculum from the Working cell bank

Inputs:

MA

Animal component–  
free medium

Qualified  
WCB

PP

post-thaw  
temp, CO<sub>2</sub>, pH

Risks

- Contamination during thawing and handling
- Recovery stress impacting cell viability and growth

Transfer Targets

- Viabilité > 95%
- Density:  
0,3-0,5 X10<sup>6</sup>  
cells/ml

Amplification

→ To bioreactor

Dish  
5 mL - 10 mL  
Day 0-1

Flask  
10 mL - 30 mL  
Day 1-3

Erlen  
30 mL - 200 mL  
Day 4-9

**Outputs :** Viable inoculum for seed train  
IPCs : viability, growth rate, morphology



# Cell Bank Thawing and Pre-culture

Preparation of a viable and stable  
inoculum from the Working cell bank

Inputs:

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Animal component–  
free medium

Qualified  
WCB

PP

post-thaw  
temp, CO<sub>2</sub>, pH

Risks

- Contamination during thawing and handling
- Recovery stress impacting cell viability and growth

Quality attributes

**Sterility / Endotoxins**  
**Cell identity (Genetic stability,**  
**Appearance / morphology,**  
**Viability)**

Dish  
5 mL - 10 mL  
Day 0-1

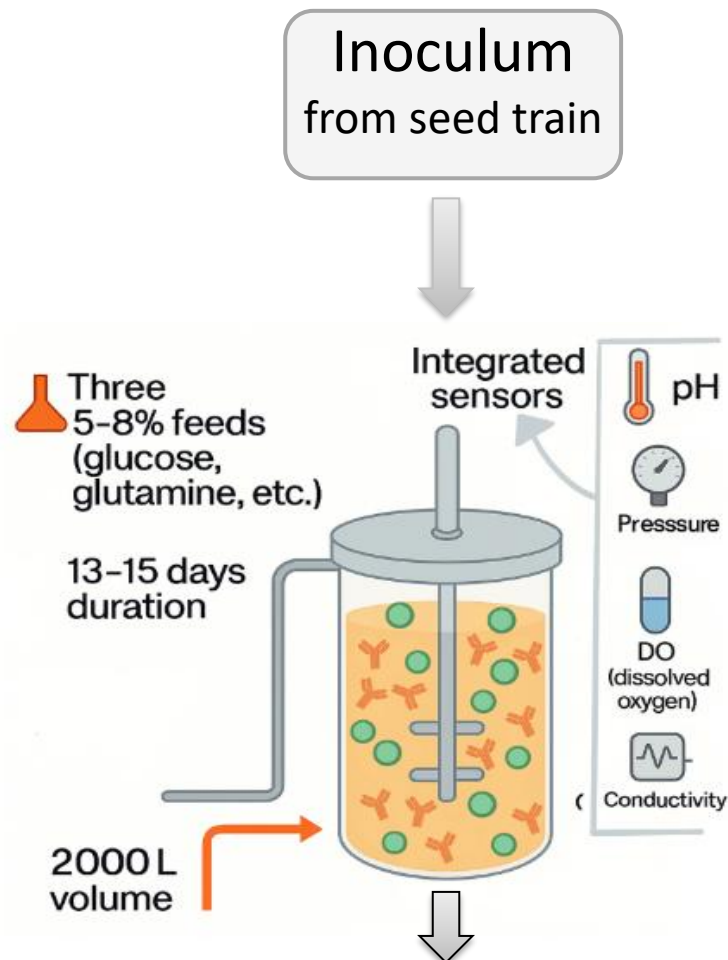
Flask  
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30 mL - 200 mL  
Day 4-9

**Outputs** : Viable inoculum for seed train  
IPCs : viability, growth rate, morphology

# Production Fed-Batch (10–14 days)

High-titer IgG synthesis



**Outputs** : Culture broth containing cells and secreted monoclonal antibody

# Production Fed-Batch (10–14 days)

High-titer IgG synthesis

Inputs:

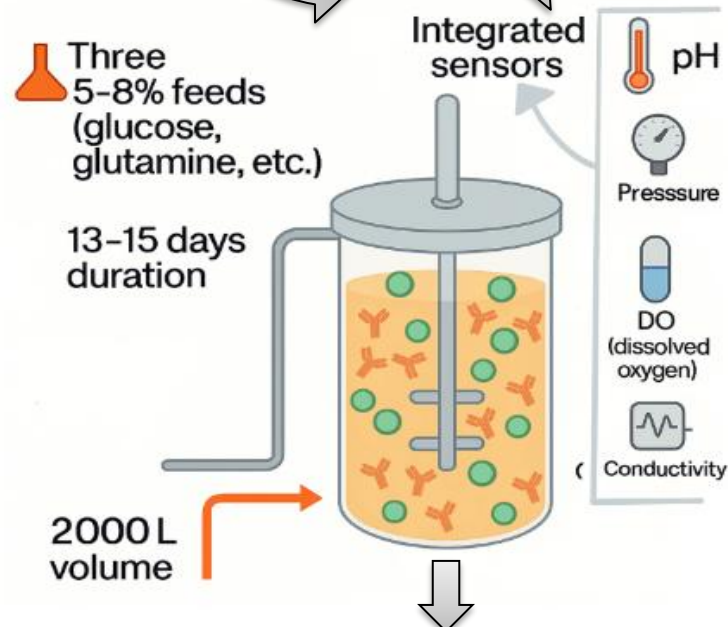
MA

Production medium  
(chemically defined)  
Feed solutions

Inoculum  
from seed train

PP

pH, dissolved oxygen,  
temperature, feeding  
rate, agitation speed



**Outputs** : Culture broth containing cells and secreted monoclonal antibody

IPCs : Viability, titer, pH, and metabolite levels

# Production Fed-Batch (10–14 days)

## High-titer IgG synthesis

### Inputs:

MA

Production medium  
(chemically defined)  
Feed solutions

Inoculum  
from seed train

PP

pH, dissolved oxygen,  
temperature, feeding  
rate, agitation speed

### Risks

- pH or oxygen deviation glycosylation
- Nutrient depletion
- cell damage

Integrated  
sensors

pH

Pressure

DO  
(dissolved  
oxygen)

Conductivity

2000 L  
volume

**Outputs** : Culture broth containing cells and secreted monoclonal antibody

IPCs : Viability, titer, pH, and metabolite levels

# Production Fed-Batch (10–14 days)

High-titer IgG synthesis

Inputs:

MA

Production medium  
(chemically defined)  
Feed solutions

Inoculum  
from seed train

PP

pH, dissolved oxygen,  
temperature, feeding  
rate, agitation speed

Risks

- pH or oxygen deviation glycosylation
- Nutrient depletion
- cell damage

Integrated  
sensors

Quality attributes

**Cell identity (Viability, appearance)**  
**Batch homogeneity / Concentration**  
**Sterility**  
**Physico-chemical properties (pH,  
osmolality, conductivity)**

2000 L  
volume

**Outputs** : Culture broth containing cells and secreted monoclonal antibody

IPCs : Viability, titer, pH, and metabolite levels

# Harvest then Clarification

Stop the culture and preserve the product (supernatant).

Cell broth  
from the bioreactor



Bioreactor

Centrifugation

harvest

depth filtration

Clarified  
supernatant

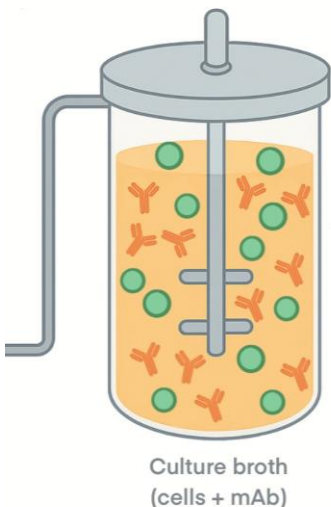


collect  
supernatant

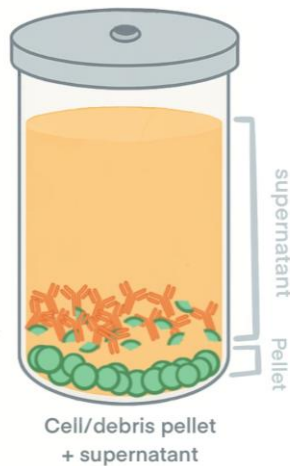


Flux (LMH)  
Pressure drop  
Media grade sequence  
Area (V/m)

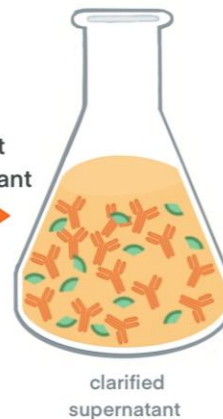
Risks  
Proteases  
Aggregates



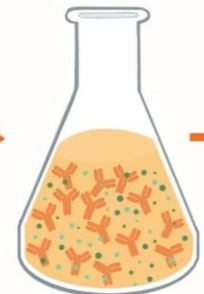
Culture broth  
(cells + mAb)



Cell/debris pellet  
+ supernatant



clarified  
supernatant



**Outputs** : Supernatant containing monoclonal antibody and cell debris

# Harvest then Clarification

Stop the culture and preserve the product (supernatant).

## Inputs:

MA

Protease inhibitor  
Antifoam agent  
Acid/base solution  
Filter material

Cell broth  
from the bioreactor

PP

pH transition, dissolved  
oxygen, temperature,  
Transfer flow rate, Hold time,  
centrifugation speed

Bioreactor

Centrifugation

harvest

Clarified  
supernatant

depth filtration

Flux (LMH)  
Pressure drop  
Media grade sequence  
Area (V/m)

Risks  
Proteases Aggregates

Stop  
culture

collect  
supernatant

clarified  
supernatant

Culture broth  
(cells + mAb)

Cell/debris pellet  
+ supernatant

supernatant  
Pellet

**Outputs** : Supernatant containing monoclonal antibody and cell debris

IPCs: Harvest timing, centrifugation parameters, transfer temperature, and clarification efficiency

# Harvest then Clarification

Stop the culture and preserve the product (supernatant).

Inputs:

MA

Protease inhibitor  
Antifoam agent  
Acid/base solution

Cell broth  
from the bioreactor

PP

pH transition, dissolved  
oxygen, temperature,  
Transfer flow rate, Hold  
time

Risks

- Protease
- Aggregation formation
- Filter clogging
- Product loss during transfer or centrifugation

harvest

collect  
supernatant

clarified  
supernatant

depth filtration

Flux (LMH)  
Pressure drop  
Media grade sequence  
Area (V/m)

Clarified  
supernatant

Risks

Proteases Aggregates

Culture broth  
(cells + mAb)

Cell/debris pellet  
+ supernatant

Pellet

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Protease inhibitor  
Antifoam agent  
Acid/base solution

Cell broth  
from the bioreactor

PP

pH transition, dissolved  
oxygen, temperature,  
Transfer flow rate, Hold  
time

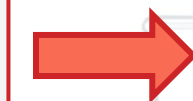
Risks

- Protease
- Aggregation formation
- Filter clogging
- Product loss during transfer or centrifugation

Quality attributes

**Identity (Intact antibody)**  
**Purity/Impurities – Aggregates**  
**Appearance / colour**

harvest



collect  
supernatant

clarified  
supernatant

Pressure drop  
Media grade sequence  
Area (V/m)

Risks

Proteases Aggregates

Culture broth  
(cells + mAb)

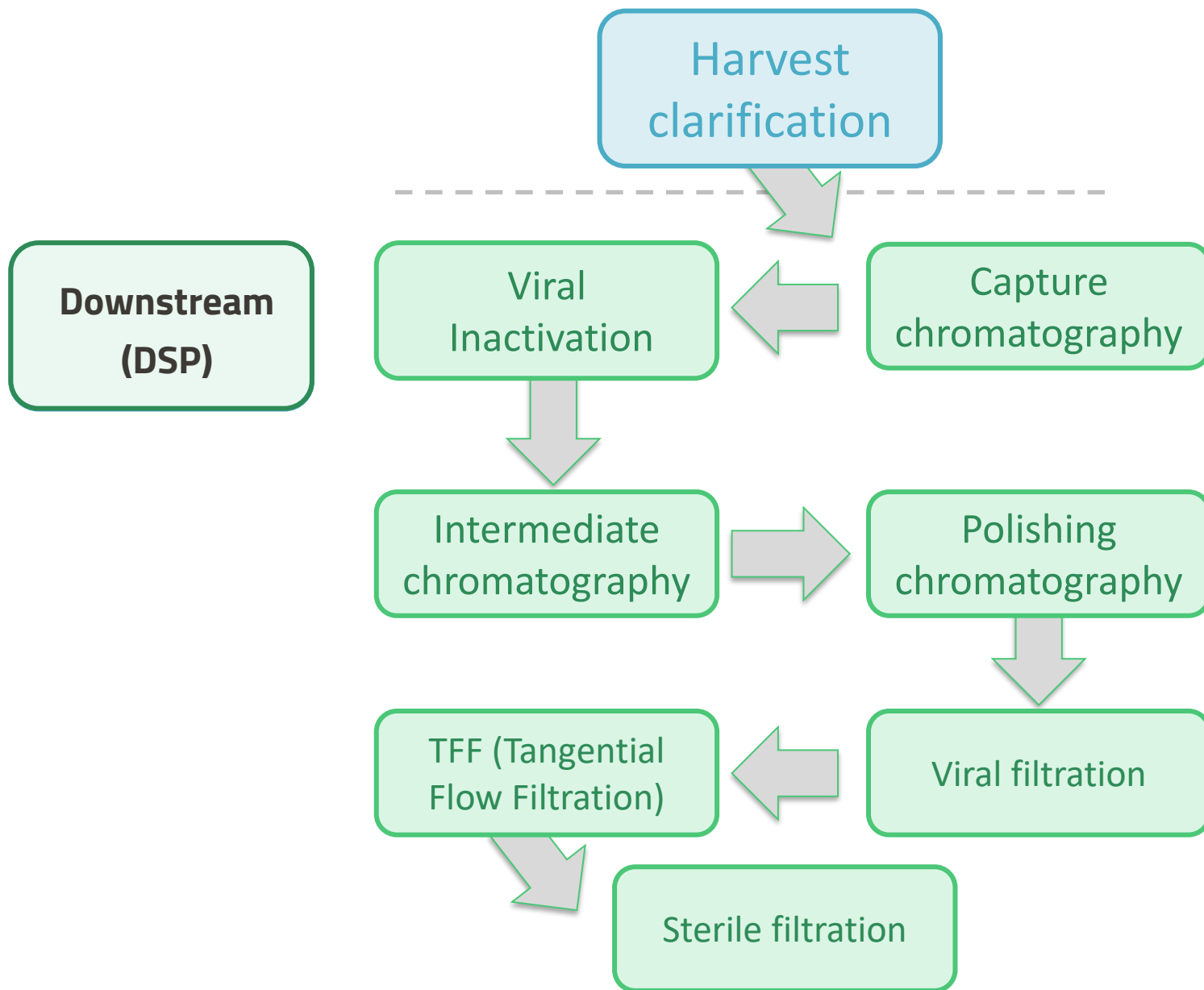
Cell/debris pellet  
+ supernatant

Pellet

**Outputs** : Supernatant containing monoclonal antibody and cell debris

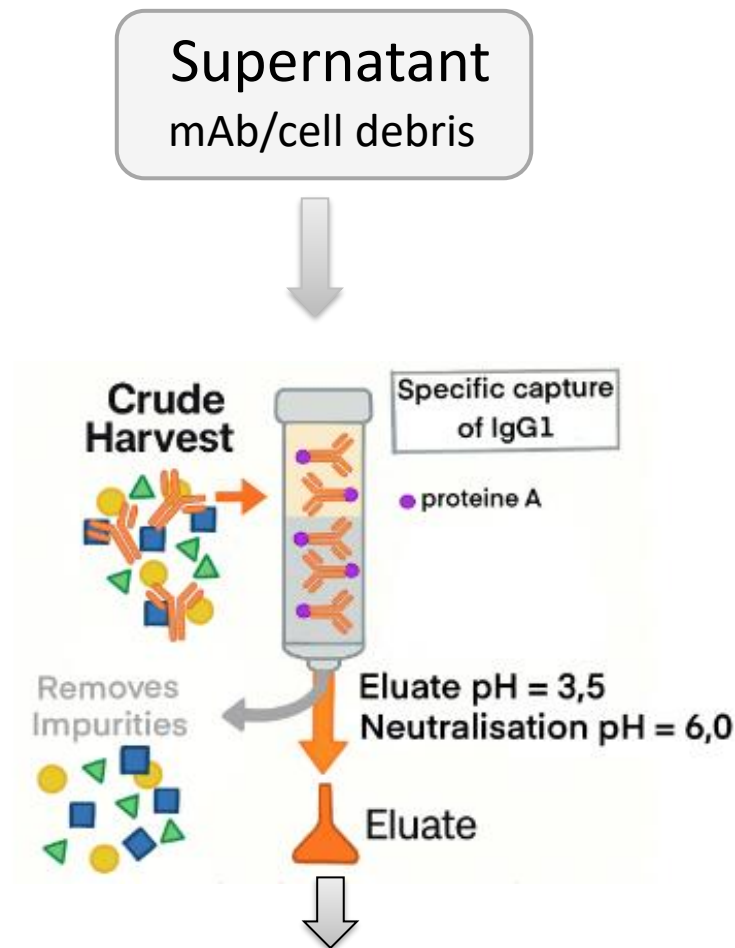
IPCs: Harvest timing, centrifugation parameters, transfer temperature, and clarification efficiency

# Downstream Process Flow



# Capture chromatography - Protein A

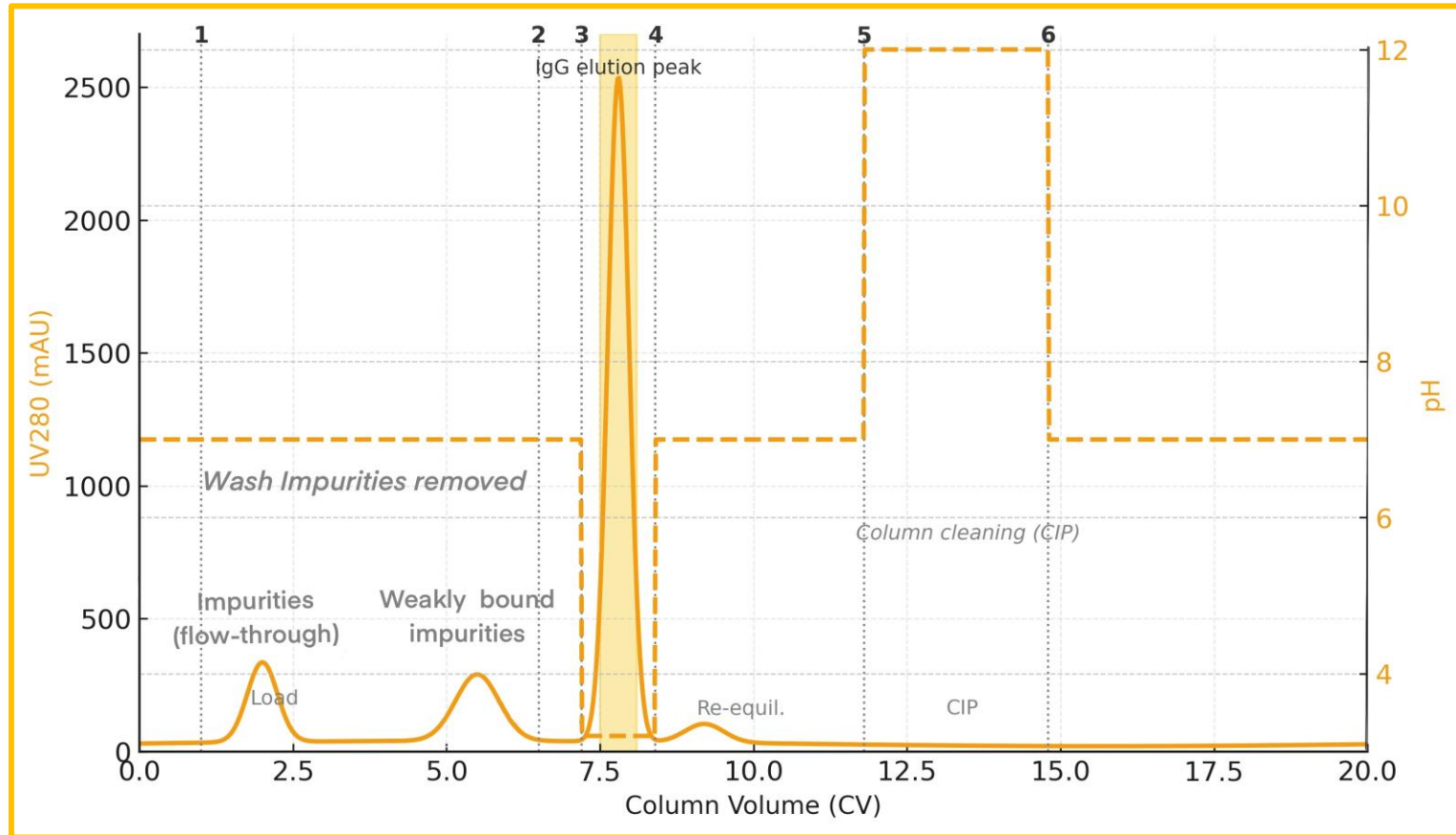
To selectively capture and purify the monoclonal antibody (IgG)



**Outputs** : Neutralized Protein A eluate containing purified monoclonal antibody

# Capture chromatography - Protein A

To selectively capture and purify the monoclonal antibody (IgG)



**Outputs :** Neutralized Protein A eluate containing purified monoclonal antibody

# Capture chromatography - Protein A

To selectively capture and purify the monoclonal antibody (IgG)

## Inputs:

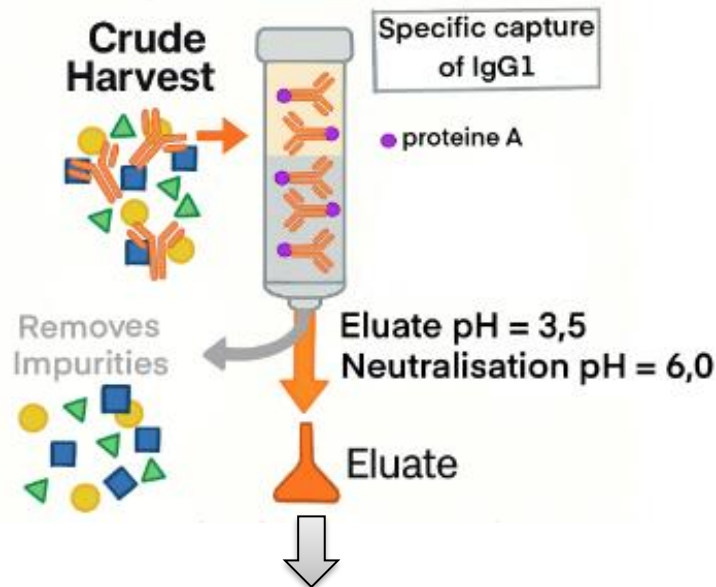
MA

Protein A resin  
Buffer pH & conductivity  
column material  
Protease inhibitors

Supernatant  
mAb/cell debris

PP

Loading density, flow rate, Wash and elution pH, and column pressure



**Outputs :** Neutralized Protein A eluate containing purified monoclonal antibody

IPCs: UV280 monitoring, eluate pH, conductivity, column pressure ( $\Delta P$ ), and protein concentration check

# Capture chromatography - Protein A

To selectively capture and purify the monoclonal antibody (IgG)

## Inputs:

MA

Protein A resin  
Buffer pH & conductivity  
column material  
Protease inhibitors

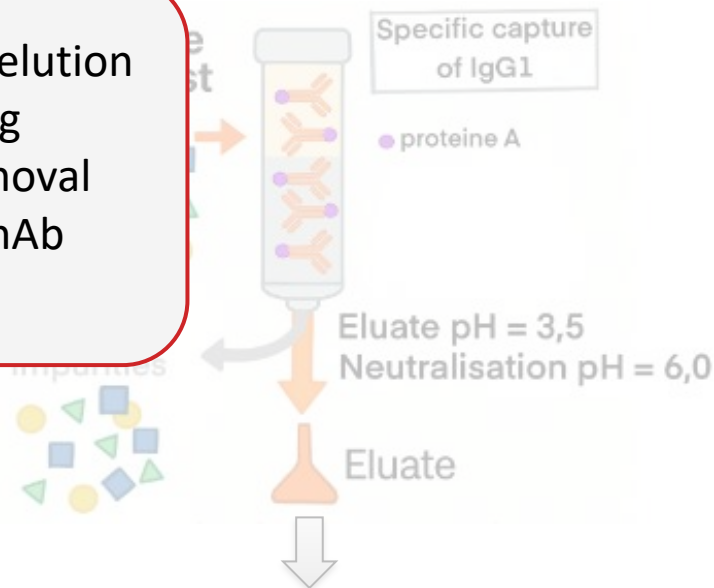
PP

Loading density, flow rate, Wash and elution pH, and column pressure

Supernatant  
mAb/cell debris

## Risks

- Aggregation during low-pH elution
- Incomplete antibody binding
- Insufficient HCP or DNA removal
- Over-acidification causing mAb denaturation



**Outputs :** Neutralized Protein A eluate containing purified monoclonal antibody

IPCs: UV280 monitoring, eluate pH, conductivity, column pressure ( $\Delta P$ ), and protein concentration check

# Capture chromatography - Protein A

To selectively capture and purify the monoclonal antibody (IgG)

## Inputs:

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Protein A resin  
Buffer pH & conductivity  
column material  
Protease inhibitors

## Risks

- Aggregation during low-pH elution
- Incomplete antibody binding
- Insufficient HCP or DNA removal
- Over-acidification causing mAb denaturation

Supernatant  
mAb/cell debris

PP

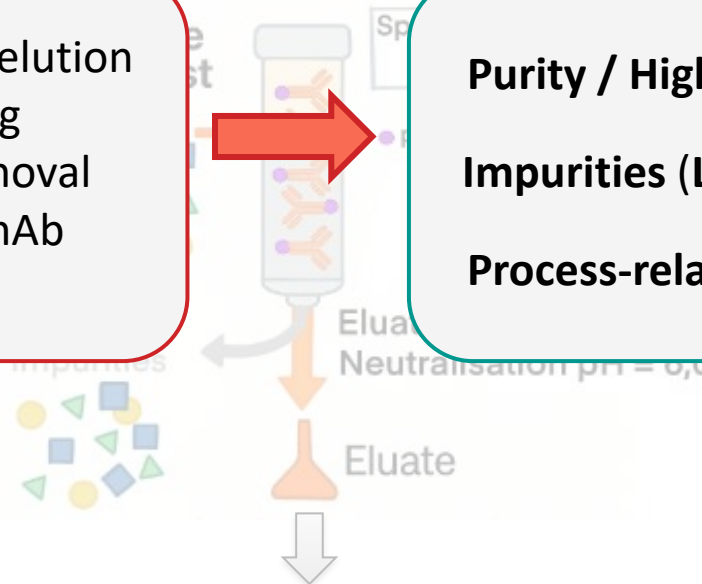
Loading density, flow rate, Wash and elution pH, and column pressure

## Quality attributes

**Purity / High recovery**

**Impurities (Low level of aggregates)**

**Process-related impurities**

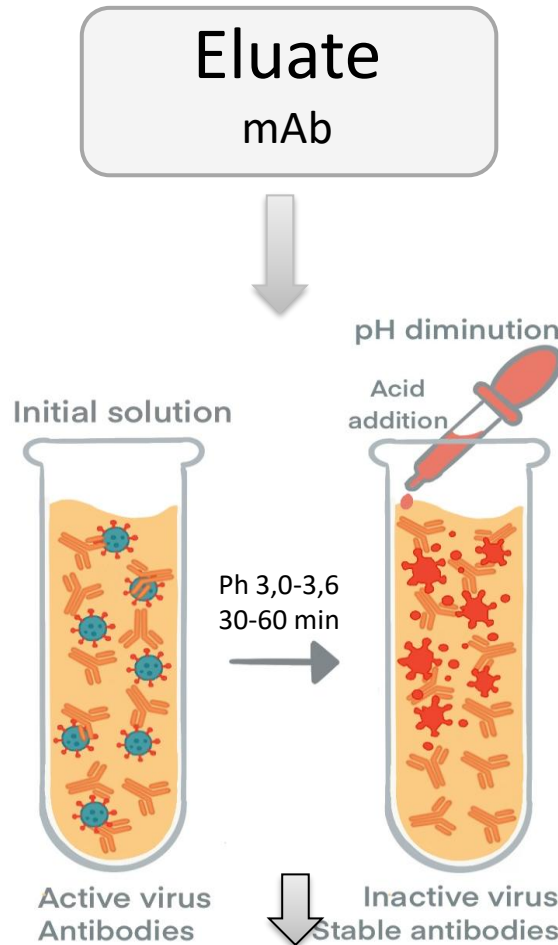


**Outputs :** Neutralized Protein A eluate containing purified monoclonal antibody

**IPCs:** UV280 monitoring, eluate pH, conductivity, column pressure ( $\Delta P$ ), and protein concentration check

# Viral inactivation (Low pH)

Inactivate enveloped viruses using an acidic pH



**Outputs** : virus-inactivated mAb pool (pH 6.0–6.5)



# Viral inactivation (Low pH)

Inactivate enveloped viruses using an acidic pH

## Inputs:

MA

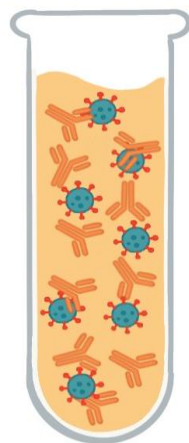
Acid reagent  
Base for neutralization  
Contact materials

Eluate  
mAb

PP

Target pH (typically 3.0–3.6), Hold time at low pH, Temperature during hold

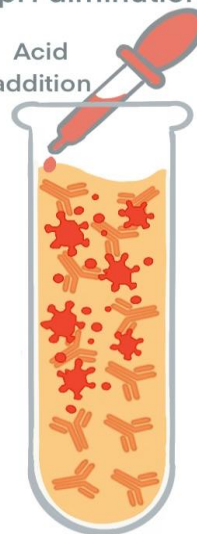
Initial solution



Active virus  
Antibodies

pH 3,0-3,6  
30-60 min

Acid  
addition



Inactive virus  
Stable antibodies

**Outputs** : virus-inactivated mAb pool (pH 6.0–6.5)

IPCs : pH setpoint, hold time, mixing uniformity (no pH gradients), neutralization

# Viral inactivation (Low pH)

Inactivate enveloped viruses using an acidic pH

## Inputs:

MA

Acid reagent  
Base for neutralization  
Contact materials

## Risks

- Antibody aggregation
- heterogeneous viral inactivation
- product degradation
- mAb instability denaturation

Eluate  
mAb

PP

Target pH (typically 3.0–3.6), Hold time at low pH, Temperature during hold

pH diminution

Acid addition

Active virus  
Antibodies

Inactive virus  
Stable antibodies

**Outputs** : virus-inactivated mAb pool (pH 6.0–6.5)

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Inactivate enveloped viruses using an acidic pH

## Inputs:

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Acid reagent  
Base for neutralization  
Contact materials

PP

Target pH (typically 3.0–3.6), Hold time at low pH, Temperature during hold

Eluate  
mAb

## Risks

- Antibody aggregation
- heterogeneous viral inactivation
- product degradation
- mAb instability denaturation

## Quality attributes

**Sterility / Endotoxins**  
**Purity / Impurities (aggregates <1%)**  
**Identity / Conformation / Stability**



pH 3,0-3,6  
30-60 min



Active virus  
Antibodies



Inactive virus  
Stable antibodies

**Outputs** : virus-inactivated mAb pool (pH 6.0–6.5)

IPCs : pH setpoint, hold time, mixing uniformity (no pH gradients), neutralization

# Polishing chromatography - CEX

Polish the mAb by removing charged impurities

## Inputs:

MA

Buffer composition  
mAb concentration  
Resin type and ligand

virus-inactivated  
mAb pool

PP

conductivity and pH  
setpoints, flow rate,  
Column pressure

Equilibration  
(Low conductivity)

Loading

typical conditions  
pH < pI of mAb;  
Conductivity 3-6 mS/cm

Elution

washing

Eluate  
impurities

mAb

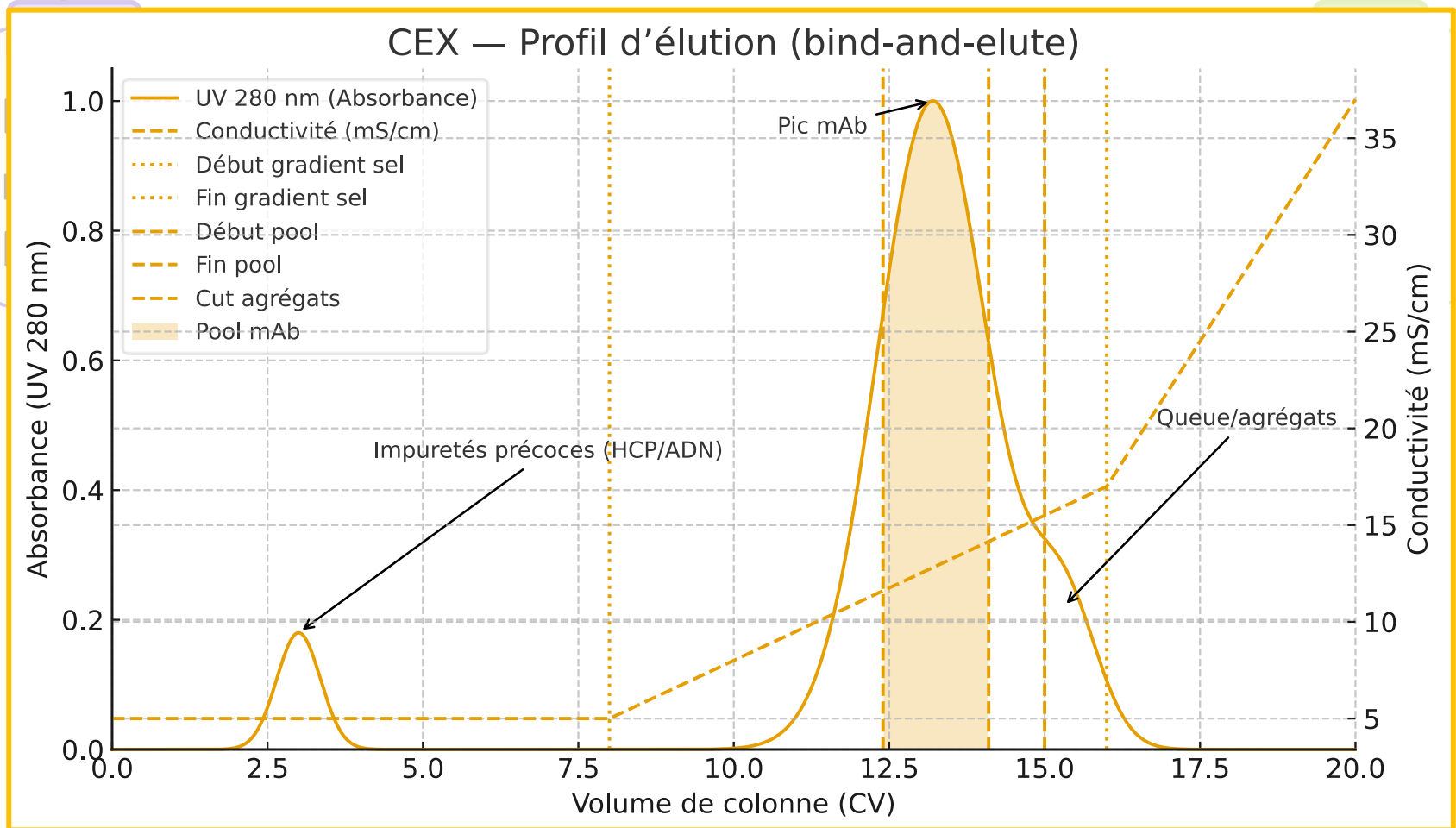
**Outputs** : Purified mAb pool – impurities removed, pH/conductivity on spec.

IPCs : conductivity and pH, UV280 monitoring, column pressure , confirmation of pool  
pH/conductivity

# Polishing chromatography - CEX

Polish the mAb by removing charged impurities

Inputs:



**Outputs :** Purified mAb pool – impurities removed, pH/conductivity on spec.

IPCs : conductivity and pH, UV280 monitoring, column pressure , confirmation of pool

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# Polishing chromatography - CEX

Polish the mAb by removing charged impurities

## Inputs:

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Buffer composition  
mAb concentration  
Resin type and ligand

virus-inactivated  
mAb pool

PP

conductivity and pH  
setpoints, flow rate,  
Column pressure

## Risks

- Poor pool selection
- Column fouling
- mAb instability denaturation

Equilibration  
(conductivity)

Loading

Elution

washing

Eluate  
impurities

mAb

pH 3,0-3,6  
30-60 min

**Outputs** : Purified mAb pool – impurities removed, pH/conductivity on spec.

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mAb pool

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conductivity and pH  
setpoints, flow rate,  
Column pressure

## Risks

- Poor pool selection
- Column fouling
- mAb instability denaturation

## Quality attributes

**Purity / Impurities**

**Purity / Impurities – Aggregates <1%**

**Identity – Conform sequence**

Equilibration  
(conductivity)

Load

ons  
D;  
cm

E

washing

Eluate  
impurities

mAb

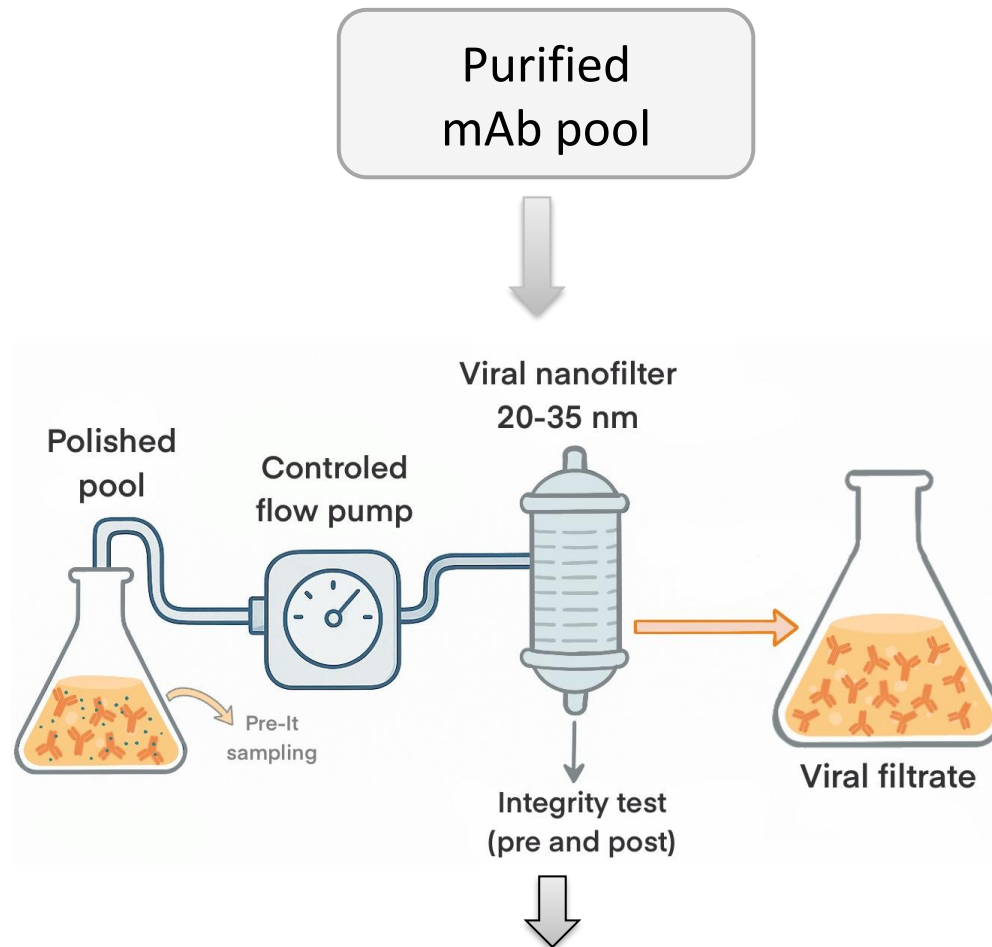
**Outputs** : Purified mAb pool – impurities removed, pH/conductivity on spec.

IPCs : conductivity and pH, UV280 monitoring, column pressure , confirmation of pool

pH/conductivity

# Viral filtration

Remove potential viral contaminants from purified mAb



**Outputs** : Purified mAb filtrate – virus and aggregates removed



# Viral filtration

Remove potential viral contaminants from purified mAb

## Inputs:

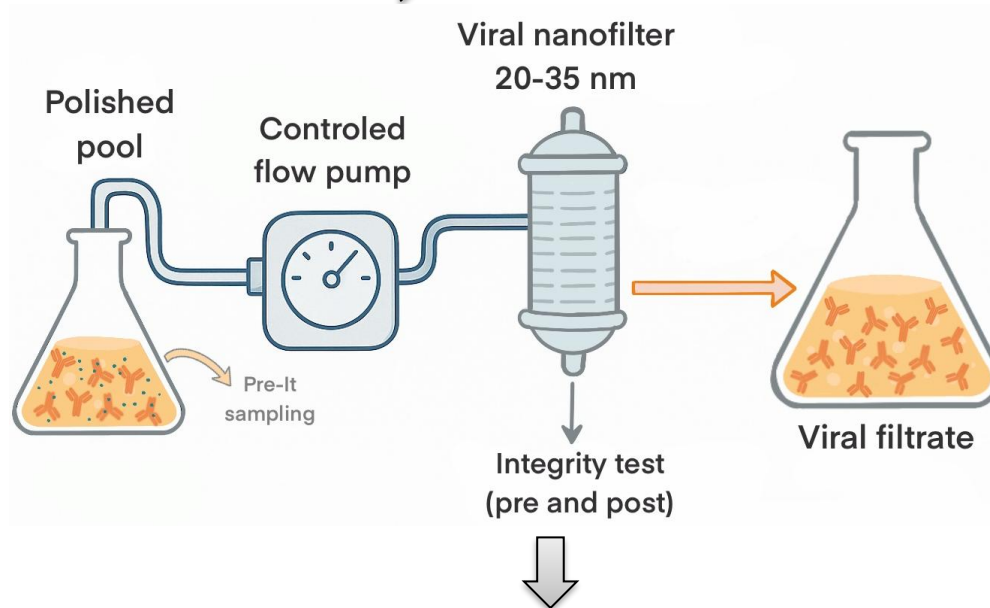
MA

Filter membrane pore size (20–35 nm)  
Buffer composition

Purified mAb pool

PP

Transmembrane pressure, flow rate, filtration time



**Outputs :** Purified mAb filtrate – virus and aggregates removed

IPCs : Flow rate monitoring, differential pressure, filter integrity (pre and post), conductivity, pH.

# Viral filtration

Remove potential viral contaminants from purified mAb

## Inputs:

MA

Filter membrane pore size (20–35 nm)  
Buffer composition

PP

Transmembrane pressure, flow rate, filtration time

Purified mAb pool

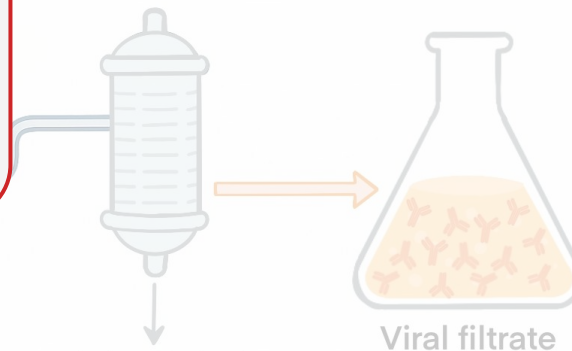
## Risks

- Filter fouling or clogging
- Filter bypass  $\Delta P$  too high
- mAb instability denaturation

Viral nanofilter  
20–35 nm



Pre-It  
sampling



Viral filtrate

Integrity test  
(pre and post)

**Outputs :** Purified mAb filtrate – virus and aggregates removed

**IPCs :** Flow rate monitoring, differential pressure, filter integrity (pre and post), conductivity, pH.

# Viral filtration

Remove potential viral contaminants from purified mAb

## Inputs:

MA

Filter membrane pore size (20–35 nm)  
Buffer composition

PP

Transmembrane pressure, flow rate, filtration time

## Risks

- Filter fouling or clogging
- Filter bypass  $\Delta P$  too high
- mAb instability denaturation

## Quality attributes

**Sterility / Endotoxins**  
**Purity / Impurities**  
**Appearance / Colour**



Viral nanofilter  
20–35 nm

Integrity test  
(pre and post)

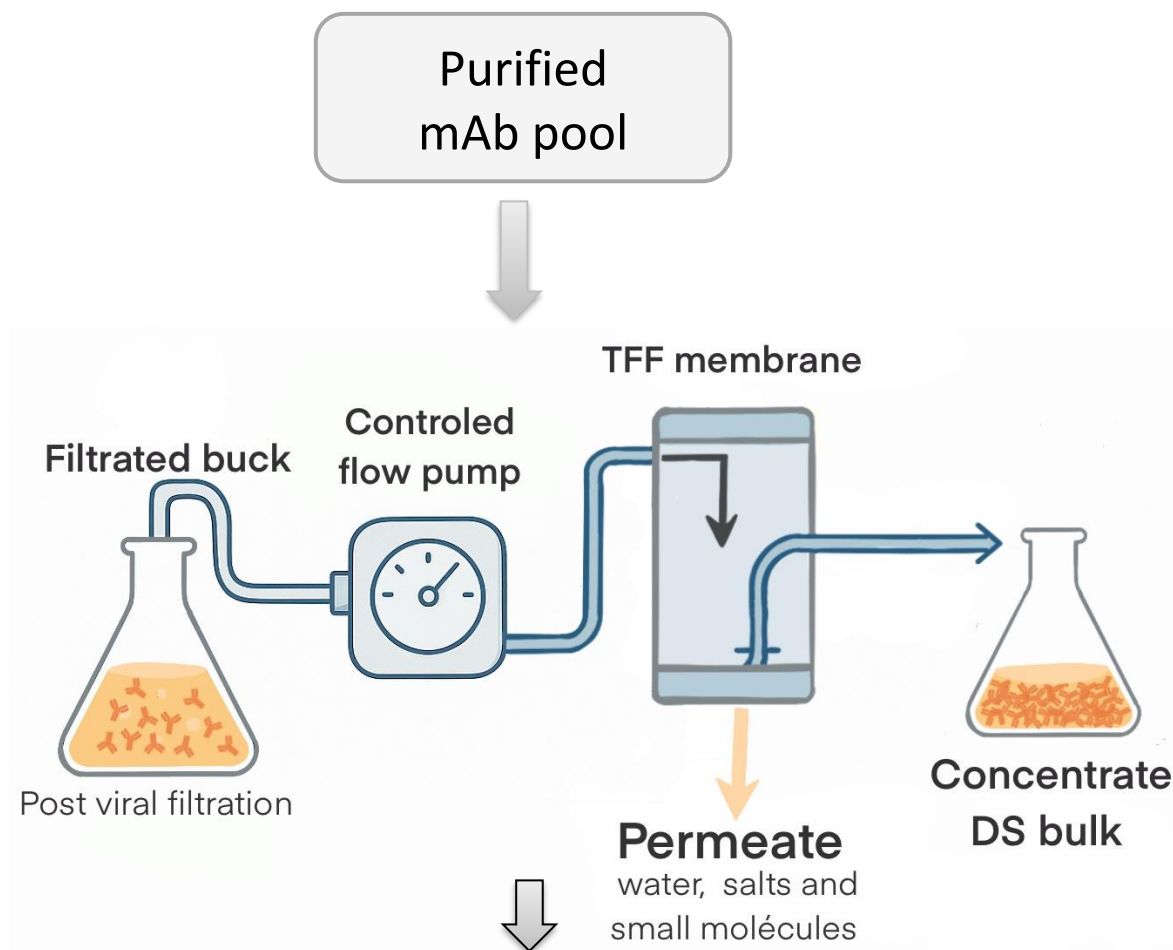
Viral filtrate

**Outputs :** Purified mAb filtrate – virus and aggregates removed

**IPCs :** Flow rate monitoring, differential pressure, filter integrity (pre and post), conductivity, pH.

# UF/DF — Tangential Flow Filtration

Concentrate the mAb and exchange into the formulation buffer



**Outputs** : concentrated mAb, buffer-exchanged to formulation buffer

# UF/DF — Tangential Flow Filtration

Concentrate the mAb and exchange into the formulation buffer

## Inputs:

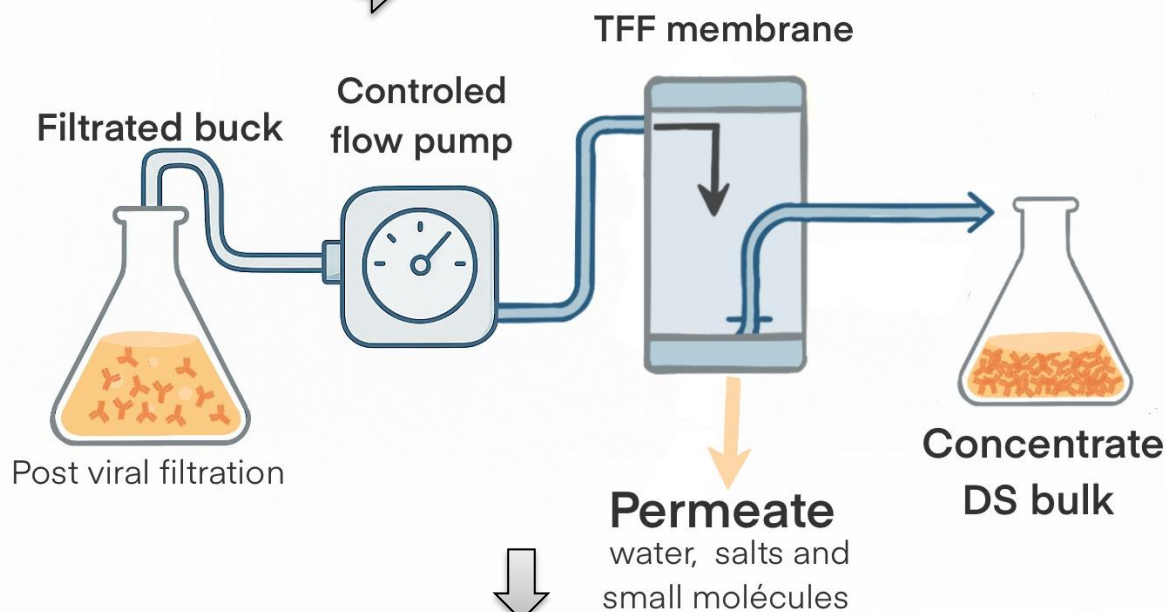
MA

TFF membrane  
Buffer composition  
Contact materials

Purified  
mAb pool

PP

TMPress setpoint &  
limits, flow rate,  
Concentration factor



**Outputs :** concentrated mAb, buffer-exchanged to formulation buffer

IPCs : UV280 monitoring, TMP, permeate flux, pH/conductivity, and turbidity check.

# UF/DF — Tangential Flow Filtration

Concentrate the mAb and exchange into the formulation buffer

## Inputs:

MA

TFF membrane  
Buffer composition  
Contact materials

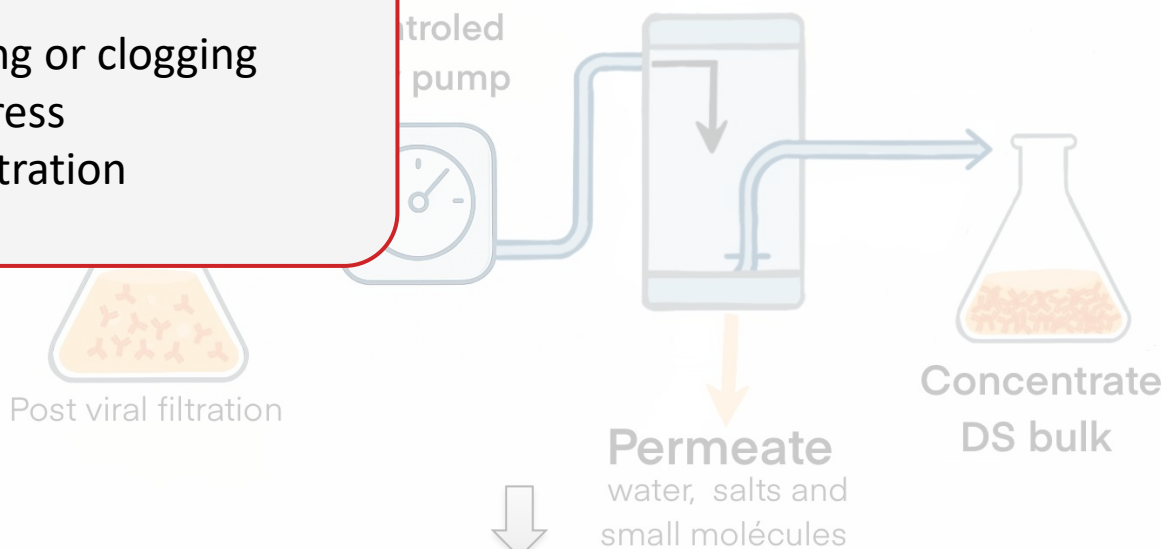
PP

TMPress setpoint &  
limits, flow rate,  
Concentration factor

Purified  
mAb pool

## Risks

- Membrane fouling or clogging
- concentration stress
- Insufficient diafiltration
- Antibodies loss



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# UF/DF — Tangential Flow Filtration

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- Insufficient diafiltration
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## Quality attributes

**Physico-chemical properties**  
**Purity / Impurities**  
**Concentration**

Post viral filtration

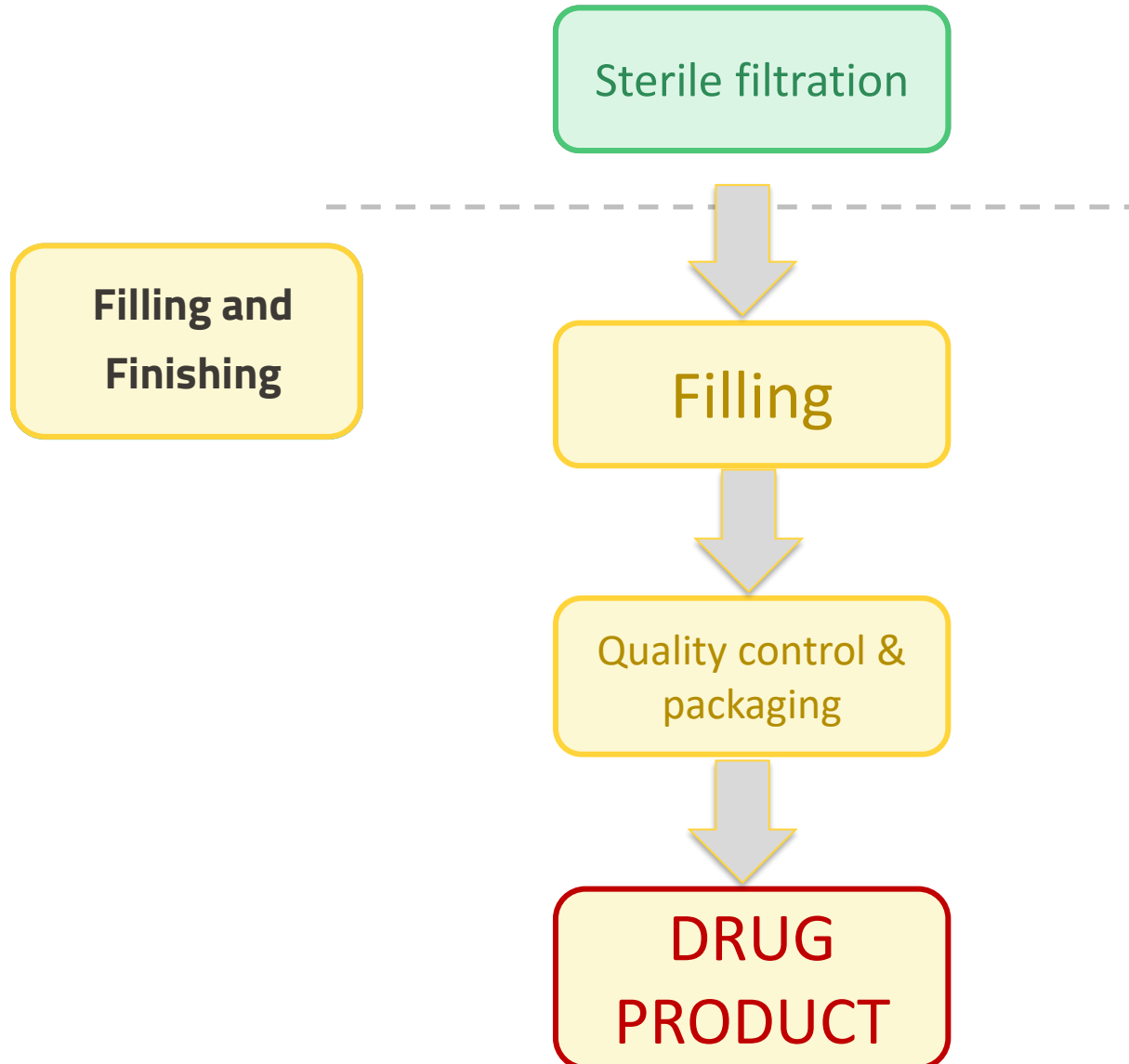
**Permeate**  
water, salts and  
small molecules

**Concentrate**  
DS bulk

**Outputs** : concentrated mAb, buffer-exchanged to formulation buffer

IPCs : UV280 monitoring, TMP, permeate flux, pH/conductivity, and turbidity check.

# Fill and Finish Process Flow





# Remplissage & Conditionnement

Perform aseptic filling, stoppering, crimping, and visual inspection

## Inputs:

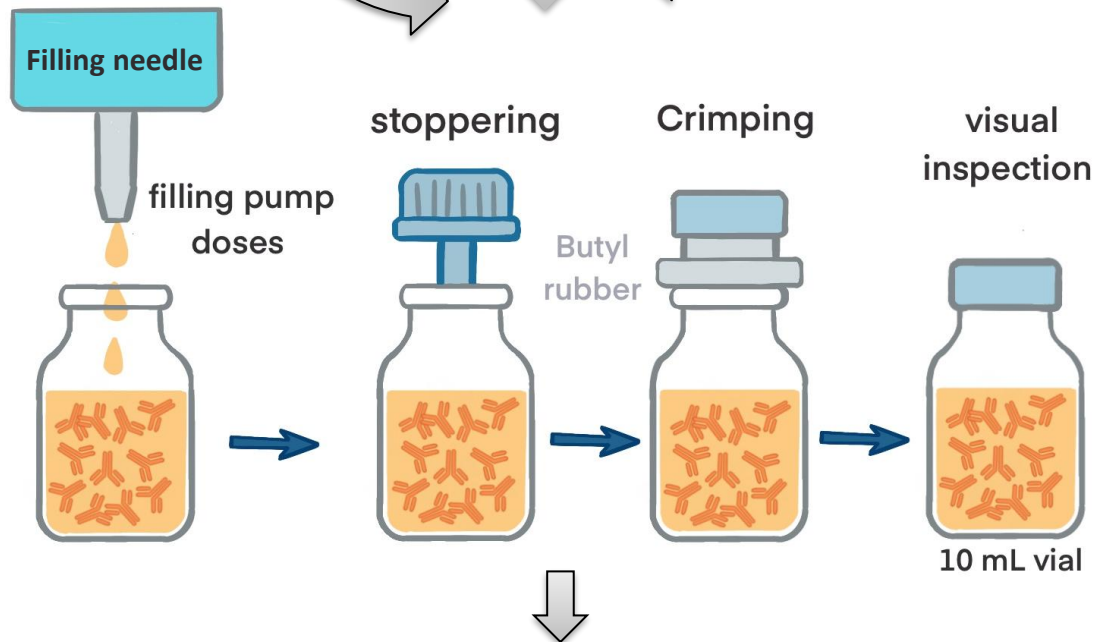
MA

TFF membrane  
Buffer composition  
Contact materials

concentrated mAb  
In formulation buffer

PP

TMPress setpoint &  
limits, flow rate,  
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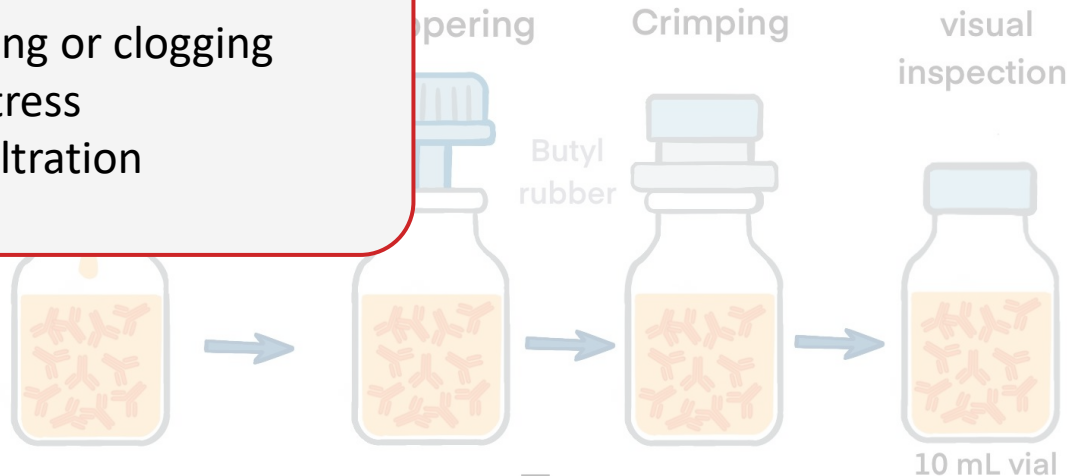
concentrated mAb  
In formulation buffer

PP

TMPress setpoint &  
limits, flow rate,  
Concentration factor

## Risks

- Membrane fouling or clogging
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concentrated mAb  
In formulation buffer

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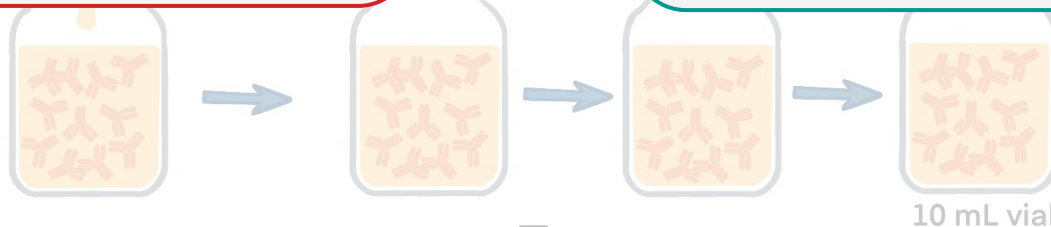
TMPress setpoint &  
limits, flow rate,  
Concentration factor

## Risks

- Membrane fouling or clogging
- concentration stress
- Insufficient diafiltration
- Antibodies loss

## Quality attributes

**Physico-chemical properties**  
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**Concentration**



**Outputs** : concentrated mAb, buffer-exchanged to formulation buffer

IPCs : UV280 monitoring, TMP, permeate flux, pH/conductivity, and turbidity check.

# Ce qui change en clinique

- Contraintes GMP/supply : hold times qualifiés, QP release, chaîne du froid
- Sécurité virale : Viral Inactivation qualifiée, filtrations virales + integrity tests
- Culture en circuit fermé
- CEX/AEX : critères de pool HCP/ADN/agrégats
- DP : media fill, intégrité fermeture, étiquetage clinique